

## COUMARONOCHROMONES FROM THE STEMS OF *EUSCHRESTA FORMOSANA*

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**Key Word Index**—*Euchresta formosana*; Leguminosae; coumaronochromone; euchretin B; euchretin C.

**Abstract**—Two new coumaronochromones, named euchretins B and C, were isolated from the stems of *Euchresta formosana* in addition to a known coumaronochromone, euchretin A. By means of the spectroscopic data, the structures of euchretins B and C were determined to be 8,3'-di( $\gamma,\gamma$ -dimethylallyl)-5,7,4',5'-tetrahydroxy- and 6,8,3'-tri( $\gamma,\gamma$ -dimethylallyl)-5,7,4',5'-tetrahydroxycoumaronochromone respectively.

### INTRODUCTION

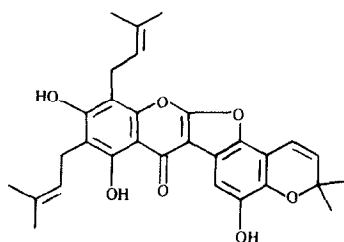
In continuation of our chemotaxonomic studies on the genus *Euchresta*, we have reported several new prenylated flavonoids from *E. japonica* [1–4]. As a result of our interest in the flavonoid compounds in *E. japonica*, our attention was drawn to the chemical constituents of *E. formosana* (Hayata) Ohwi because the species has been hypothesized to evolve from proto-*Euchresta* by a different pathway from *E. japonica* [5]. It has been used in folk medicines as a pain-killer, particularly for the throat and for snake wounds in Taiwan [6]. Our present investigation on the constituents of the stems of this plant resulted in the isolation of two novel coumaronochromones. The structure elucidation of which is described in this paper.

### RESULTS AND DISCUSSION

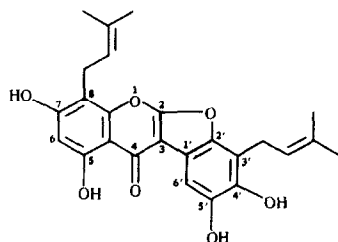
Compound 1, mp 208–209°,  $C_{30}H_{36}O_7$  ( $M^+$ :  $m/z$  502.1992, calcd. 502.1991), was identified as euchretin A. This was previously isolated from the stems of *E. japonica* [3].

Compound 2, mp 242–243°,  $C_{25}H_{24}O_7$  ( $M^+$ :  $m/z$  436.1501, calcd. 436.1521), was also obtained as a colourless powder. The  $^1H$  NMR spectrum contained, two two-proton doublets ( $J = 7.3$  Hz) at  $\delta$  3.52 and 3.62, two one-proton triplets ( $J = 7.3$  Hz) at  $\delta$  5.26 and 5.38 and four

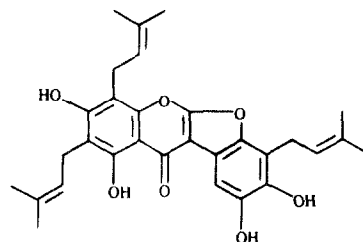
methyl groups at  $\delta$  1.68, 1.70, 1.86 and 1.89. This suggested that 2 had two  $\gamma,\gamma$ -dimethylallyl groups. Furthermore, four one-proton singlets at  $\delta$  7.53, 8.77, 9.69 and 12.59 showed the presence of four hydroxyl groups. Each absorption band in the IR (3450 and 1650  $cm^{-1}$ ; chelated OH and  $>C=O$ ) and the UV (260, 288, 308sh and 348 nm) spectra indicated 2 was an isoflavone derivative. A coumaronochromone structure was indicated by the absence of a characteristic signal due to a proton at C-2 (ca 8 ppm) in the  $^1H$  NMR spectrum. Two aromatic protons appeared at  $\delta$  6.40 and 7.34. The latter signal was assigned to a proton at C-6' which was shielded diamagnetically by an adjacent carbonyl group. The former was assigned to a proton on the A ring. By comparison with the known structure of euchretin A (1), the four hydroxyl groups were assigned at C-5, 7, 4' and 5', and one of two  $\gamma,\gamma$ -dimethylallyl groups was placed at C-3'. Hence the other  $\gamma,\gamma$ -dimethylallyl group had to be at either C-6 or C-8. In the  $^{13}C$  NMR of the permethyl ether of 2, four methoxyl carbons were observed at  $\delta$  55.99, 56.20, 56.66 and 61.31. The last signal in the low-field was assigned to C-4'. This finding suggested that all the methoxy groups, except C-4', were not sandwiched between any substituents. Hence the position of the  $\gamma,\gamma$ -dimethylallyl group on the A ring was deduced to be at C-8. Consequently, the structure of 2 was concluded to be 8,3'-di( $\gamma,\gamma$ -dimethylallyl)-5,7,4',5'-tetrahydroxy coumaronochromone, which we have named euchretin B.



1



2



3

Compound **3**, mp 215° (dec.),  $C_{30}H_{32}O_7$  ( $M^+$ :  $m/z$  504.2146, calcd. 504.2148), was obtained as colourless needles. All the spectral data were similar to those of **1** and **2**. These results suggested that **3** had a coumaronochromone skeleton. In the  $^1H$  NMR, four hydroxyl groups at  $\delta$  7.56, 8.34, 8.81 and 13.33, three  $\gamma,\gamma$ -dimethylallyl groups [ $\delta$  1.67, 1.69, 1.70, 1.79, 1.86 and 1.89 (3H, s, Me), 3.45 (2H,  $d$ ,  $J=6.9$  Hz  $CH_2$ ), 3.59 (4H,  $m$ ,  $2 \times CH_2$ ), 5.22–5.39 (3H,  $m$ ,  $3 \times CH=C<$ )] and only a single aromatic proton ( $\delta$  7.35) assignable to H-6', were observed. The UV spectra data suggested that **3** had the same oxygenation pattern as **1** and **2**. Therefore, it could be concluded that **3** had the same partial structure as **1** about the A ring moiety and had the same one as **2** about the B ring. Consequently, the structure of **3** was confirmed to be 6,8,3'-tri( $\gamma,\gamma$ -dimethylallyl)-5,7,4',5'-tetrahydroxycoumaronochromone, which we have named euchretin C. A facile distinction of **2** and **3** from other compounds is provided by the observation that each UV band I undergoes an abnormal bathochromic shift to 580–590 nm upon addition of sodium methoxide, due to the formation of *ortho* quinone.

Other flavonoid compounds in the stems as well as the roots of *E. formosana* are now being investigated.

#### EXPERIMENTAL

**Plant material.** The stems of *Euchresta formosana* were collected in Taipei (Taiwan) in Aug. 1987. Voucher specimens are deposited at the Herbarium of Gifu Pharmaceutical University.

**Extraction and isolation of compounds 1–3.** Dried stems (250 g) of *E. formosana* were crushed into pieces and extracted with  $CH_2Cl_2$ ,  $Me_2CO$  and MeOH, successively. The  $CH_2Cl_2$  soln was concentrated *in vacuo* to give a greenish-gum (12 g), which was subjected to silica gel CC. Fractions eluted *n*-hexane– $Me_2CO$  (5:1) contained a mixture of **1**–**3**. The mixture was further purified by CC (*n*-hexane– $Me_2CO$ ) and the separated compounds were recrystallized to give **1** (5 mg), **2** (25 mg) and **3** (32 mg).

**Euchretin B (2).**  $C_{25}H_{24}O_7$ ,  $M_r$  436.1501 (calcd 436.1521), a colourless powder ( $CHCl_3$ – $Et_2O$ ), mp 208–209°. EIMS ( $m/z$ ) (rel. int.): 436 (95) [ $M$ ] $^+$ , 421 (7) [ $M-Me$ ] $^+$ , 419 (13), 393 (5) [ $M-C_3H_7$ ] $^+$ , 380 (100), 365 (60), 337 (7), 325 (30), 312 (37), 203 (17), 189 (13), 165 (13); IR ( $\nu^{KBr}$   $cm^{-1}$ ): 3450, 1650, 1610, 1595; UV  $\lambda^{MeOH}$  nm (log  $\epsilon$ ): 260 (4.5), 288 (4.2), 308sh (4.0), 348 (4.0);

+ NaOMe 270, 315, 356, 580 (dec.); +  $AlCl_3$  273, 304, 370, 398; +  $AlCl_3/HCl$  269, 280sh, 294sh, 390; + NaOAc 253, 294sh, 308sh, 324sh; + NaOAc/ $H_3BO_3$  263, 292, 305, 356;  $^1H$  NMR ( $Me_2CO-d_6$ , 270 MHz):  $\delta$  1.68, 1.70, 1.86, 1.89 (3H, each  $br$ , Me), 3.52, 3.62 (2H, each  $br$   $d$ ,  $J=7.3$  Hz,  $CH_2CH=C<$ ), 5.26, 5.38 (1H, each  $br$   $t$ ,  $J=7.3$  Hz,  $CH=C<$ ), 6.40 (1H, s, H-6), 7.34 (1H, s, H-6'), 7.53, 8.77, 9.69 (1H, each s, OH), 12.95 (1H, s,  $C_5$ -OH). **Euchretin B permethylether: 2** (15 mg) was methylated with  $MeI-K_2CO_3$  in  $Me_2CO$  to give the permethyl ether (12 mg), colourless needles (MeOH), mp 197–198° (MeOH). EIMS ( $m/z$ ) (rel. int): 492 [ $M$ ] $^+$  (100:  $C_{29}H_{32}O_7$ );  $^1H$  NMR ( $CDCl_3$ , 270 MHz):  $\delta$  1.70 (6H,  $br$  s,  $2 \times$  Me), 1.86, 1.87 (3H, each  $br$  s, Me), 3.55, 3.61 (2H, each  $br$   $d$ ,  $J=7.3$  Hz,  $CH_2C=C<$ ), 3.86, 3.94, 3.97, 4.02 (3H, each s, OMe), 5.20, 5.32 (1H, each  $br$   $t$ ,  $J=7.3$  Hz,  $CH=C<$ ), 6.49 (1H, s, H-6), 7.56 (1H, s, H-6');  $^{13}C$  NMR ( $CDCl_3$ ):  $\delta$  17.83, 17.87, 25.78, 15.85 (Me), 22.06, 23.48 ( $CH_2$ ), 121.58, 121.63 ( $CH=C<$ ), 132.35, 132.60 ( $CH=C<$ ), 174.5 ( $>C=O$ ), 55.99, 56.20, 56.55, 61.31 (OMe).

**Euchretin C (3).**  $C_{30}H_{32}O_7$ ,  $M_r$  504.2146 (Calcd 504.2148), colourless needles ( $C_6H_6$ –*n*-hexane), mp 215° (dec.). EIMS ( $m/z$ ) (rel. int.): 504 (92) [ $M$ ] $^+$ , 461 (74), 449 (58), 433 (21), 407 (39), 393 (100), 377 (13), 349 (19), 337 (22), 189 (16); IR ( $\nu^{KBr}$   $cm^{-1}$ ): 3450, 1645, 1610. UV  $\lambda^{MeOH}$  nm: 263, 288, 350; + NaOMe 262, 278, 310, 370, 590 (dec.); +  $AlCl_3$  265, 295, 318sh, 349; +  $AlCl_3/HCl$  264, 288, 308sh, 349; + NaOAc 264, 290, 305sh, 354; + NaOAc/ $H_3BO_3$  267, 293, 313sh, 365.  $^1H$  NMR ( $Me_2CO-d_6$ ):  $\delta$  1.67, 1.69, 1.70, 1.79, 1.86, 1.89 (3H, each  $br$  s, Me), 3.45 (2H,  $br$   $d$ ,  $J=6.9$  Hz,  $CH_2$ ), 3.59 (4H,  $m$ ,  $2 \times CH_2$ ), 5.22–5.39 (3H,  $m$ ,  $3 \times CH=C<$ ), 7.35 (1H, s, H-6'), 7.56, 8.34, 8.81 (1H, s, OH), 13.33 (1H, s,  $C_5$ -OH).

#### REFERENCES

- Mizuno, M., Tamura, K., Tanaka, T. and Iinuma, M. (1988) *Phytochemistry* **27**, 1831.
- Mizuno, M., Tamura, K., Tanaka, T. and Iinuma, M. (1988) *Phytochemistry* **27**, 2975.
- Mizuno, M., Tamura, K., Tanaka, T. and Iinuma, M. (1988) *Heterocycles* **27**, 2047.
- Mizuno, M., Tamura, K., Tanaka, T. and Iinuma, M. (1989) *Chem. Pharm. Bull.* **37**, 195.
- Kan, W.-S. (1970) *Manual of Medicinal Plants in Taiwan* **2**, p. 290. National Research Institute of Chinese Medicine, Taiwan.
- Ohashi, H. and Sohma, K. (1970) *J. Fac. Sci. Tokyo, III* **10**, 207.